	-						
Semester	Sem III & IV (22+22 Credits)						
Subject	Theory			Lab			Workload
	Course	Credit	hours	Course	Credit	hours	
Foundation	1	2	2	-	-	-	2
Elective	1	2	2	-	-	-	2
Core 1	3	2+2+2	2+2+2	1	3	6	6 Th +
	5	2+2+2	2+2+2	T	5	D	6hr (per batch)
Core 2	2	2+2+2	2.2.2	1	n	c	6 Th +
	3	2+2+2	2+2+2	L	3	6	4hr (per batch)
Total	8	16	16	2	6	12	22+22

Programme: M. Sc. Integrated Biotechnology

Semester-III

Core 1

- Course 1: Instrumentation & Techniques-I
- Course 2: Applied Microbiology
- Course 3: Fundamentals of Immunology

Core 2

- Course 1: Plant Physiology-I
- Course 2: Animal Physiology-I

Course 3: Microbial Physiology-I

<u>Practical Core 1:</u> Total Experiments-09 (3 from each course) <u>**Practical Core 2:**</u> Total Experiments-09 (3 from each course)

Semester-IV

<u>Core 1</u>

- Course 1: Instrumentation & Techniques-II
- Course 2: Enzymes & Coenzymes
- Course 3: Introduction to Molecular Biology

<u>Core 2</u>

- Course 1: Plant Physiology-II
- Course 2: Animal Physiology-II
- Course 3: Microbial Physiology-II

<u>Practical Core 1:</u> Total Experiments-09 (3 from each course) <u>Practical Core 2:</u> Total Experiments-09 (3 from each course)

CORE: 1; Course: 01 INSTRUMENTATION & TECHNIQUES-I

UNIT –1

• **Potentiometry :** pH Electrodes : reference electrode, glass electrode, combined electrode, construction and use of pH meter, its operation, Maintenance of electrodes, ion selective meter and electrode for Ca⁺², F, Biomembrane electrode.

UNIT –2

- **Microscopy:** Phase contrast microscopy: its principle, instrumentation and application, Dark field microscopy: Principle, instrumentation and Application, Florescence microscopy: its principle instrumentation and application.
- Radio isotopes techniques :
 - Detection & measurement and use of radio activity in Biology
 - Geiger-Muller Counter: Working principle and application
 - Scintillation counter: Working principle and application
 - Autoradiography: Technique and applications

UNIT – 3

- Centrifugation :
 - Basic principles of sedimentation,
 - o types of centrifuges,
 - types of rotors,
 - Separation methods in preparative ultracentrifuges
 - Differential centrifugation
 - density gradient centrifugation
 - Analysis of subcellular fractions
 - Applications of Analytical ultracentrifuge
 - Determination of relative molecular Mass
 - Estimation of purity of macromolecules
 - Conformational changes in macromolecules
 - Safety aspects in use of centrifuge.

UNIT-4

• Spectrophotometry:

- Molecular absorption spectroscopy
 - Laws of photometry
 - Colorimetry: Components of the instruments, Applications
 - Spectrophotometer: Instrument components- single beam and double beam instrument, -Applications
 - Quantitative analysis by spectrophotometer -manual and automated
 - Fluorescence and phosphorence (principle and application)

REFERENCES:

- 1. Keith Wilson & John Walker (Ed.) (2000): Practical Biochemistry Principles & Techniques. Cambridge University Press.
- 2. Skoog, Holler and Nieman, Instrumental analysis Saunders college publication
- 3. Skoog, West and Holler, Fundamentals of Analytical Chemistry Saunders college publication
- 4. James S. Fritz & George H. Schenk, Jr. (1969): Quantitative Analytical Chemistry (2nd edition). Allyn & Bacon, Inc., Boston.
- 5. Brown S.B. (1980): An Introduction to Spectroscopy for biochemists. Academic Press London.
- 6. E.D.P. De. Robertis & E.M.F. De Robertis Jr. (2001) : Cell and Molecular Biology (8th edn.) Lippincott Williams & Wilkins, London.

CORE: 1; Course: 02 APPLIED MICROBIOLOGY

UNIT 1: MICROBIOLOGY OF WATER

- 1.1 Water purification and sanitary analysis
 - 1.1.1 Sanitary analysis of waters
- 1.2 Wastewater Treatment
 - 1.2.1 Measuring water quality
 - 1.2.2 Wastewater Treatment Process
 - 1.2.3 Home treatment systems

UNIT 2: MICROBIOLOGY OF FOOD

- 2.1 Microorganism Growth in Foods
- 2.2 Microbial Growth and Food Spoilage
- 2.3 Controlling Food Spoilage
- 2.4 Food-borne Diseases
- 2.5 Detection of food-borne pathogens

UNIT 3: MICROBIOLOGY OF FERMENTED FOOD

- 3.1 Microorganism Growth in Foods
- 3.2 Fermented milks
- 3.3 Cheese production
- 3.4 Meat and fish
- 3.5 Production of alcoholic beverages
- 3.6 Production of breads
- 3.7 Other fermented foods

UNIT 4: NEWER PROCESSES

- 4.1 Biodegradation and bioremediation processes
- 4.2 Stimulating biodegradation
- 4.3 Bioaugmentation
- 4.4 Biosensors
- 4.5 Biopesticides

REFERENCES:

1. Willey, J.M., Sherwood, L. M., & Woolverton, C. J., (2008). *Prescott, Harley & Klein's Microbiology*, 7Ed, The McGraw-Hill Companies, Inc.

CORE: 1; Course: 03 FUNDAMENALS OF IMMUNOLOGY

UNIT 1: INTRODUCTION TO IMMUNE SYSTEM

- 1.1 Overview of host resistance
- 1.2 Cells of the immune system
- 1.3 Organs and Tissues of the immune system
- 1.4 Physical barriers in non-specific resistance

UNIT 2: NON-SPECIFIC HOST RESISTANCE

- 2.1 Phagocytosis
- 2.2 Inflammation
- 2.3 Chemical mediators in non-specific (Innate) resistance
 - 2.3.1 Antimicrobial peptides
 - 2.3.2 Complement
 - 2.3.3 Cytokines
 - 2.3.4 Acute-Phase proteins

UNIT 3: SPECIFIC HOST RESISTANCE-I

- 3.1 Overview of specific immunity
- 3.2 Antigens
- 3.3 Types of specific immunity
- 3.4 Recognition of Foreignness
- 3.5 T cell Biology

UNIT 4: SPECIFIC HOST RESISTANCE-II

- 4.1 B cell Biology
- 4.2 Immunoglobulin Structure
- 4.3 Immunoglobulin Function
- 4.4 Immunoglobulin Classes
- 4.5 Antibody kinetics
- 4.6 Action of Antibodies

REFERENCES:

1. Willey, J.M., Sherwood, L. M., & Woolverton, C. J., (2008). *Prescott, Harley & Klein's Microbiology*, 7Ed, The McGraw-Hill Companies, Inc.

CORE: 2; Course: 01 PLANT PHYSIOLOGY-I

UNIT – 1 Plant-water relations:

Water relations- Diffusion, Diffusion pressure deficit, Osmosis, types, Osmotic pressure, Turgor pressure, Wall pressure and interrelationship, Solute potential, water potential.

Water absorption-mechanism, factors affecting rate of water absorption.

Transpiration -Definition, types, Significance.

Mechanism of opening and closing of stomata- concept of pH in opening and closing of stomata, Theory of proton transport, stomatal opening in succulent plants, factors influencing transpiration rate, Antitranspirants.

UNIT -2 Ascent of Sap and Organic translocation:

Vital theories, Root pressure theories, Cohesion and adhesion, Physical force theories Merits and demerits of theories.

Electrophysiology- Action potential in plants.

Organic Translocation: Phloem sap, Phloem loading and unloading, Mass-flow (pressure flow) hypothesis- Critical evaluation.

UNIT-3 Physiology of flowering:

Concept of photomorphogenesis -Physiology of flowering, Theories related to flowering, Role of light, Phytochrome system, role and mechanism of action, Florigen concept. Photoperiodism and plant types, Vernalization.

UNIT -4 Growth and development:

Mineral nutrition: Macro, and micronutrients, their role, deficiency.

Growth: general aspects, Pattern and stages of growth, phases of growth, factors affecting growth. Types of plant movements: Tactic, tropic and nastic movements.

REFERENCES:

- 1. Frank B.Salisbury and Cleon W.Ross (2010), Plant Physiology, Cengage learning products, India Edition.
- 2. S.K. Verma and Mohit Verma (1999) Plant Physiology Biochemistry and Biotechnology, S Chand.
- 3. Lincoln Taiz and Eduardo Zaiger (Fourth Edition), Plant Physiology, Sinquur Associates inc. Publishers.
- 4. S.N. Pandey ajd K.K Singh, Plant physiology, Vikas Pub.

CORE: 2; Course: 02 ANIMAL PHYSIOLOGY-I

UNIT-1 Tissues and Glands

(Ch 4, Tortora & Grabowski)

(Ch 64-66, Guyton & Hall)

Concepts and classification- Epithelial tissue, Connective tissue, Muscular tissue, Nervous tissue and Types of glands.

UNIT-2 Digestive System

Histology and functions of gastrointestinal tract and its associated glands; Mechanical and chemical digestion of food; Role of gastrointestinal hormones; Control and action of GI Tract secretions; Absorptions of carbohydrates, lipids, proteins, water, minerals and vitamins; Disorders of the digestive system.

UNIT-3 Nervous System

(Ch 12, 17, Tortora & Grabowski)

General organization: Neuron resting membrane potential and its basis; Origin of action potential and its propagation in myelinated and unmyelinated nerve fibers; Synaptic transmission and types of synapsis, Neuro-muscular junction; Reflex activity-reflex arc; Types of reflexes, Physiology of hearing and vision.

UNIT 4 Muscle

(Ch 6, 7, Guyton & Hall/ Ch 10, Tortora & Grabowski)

Histology of different types of muscle; Ultra structure of skeletal muscle; Molecular and chemical basis of muscle contraction; Characteristics of muscle twitch; Motor unit, summation, tetanus and muscle dystrophies.

CORE: 2; Course: 03 MICROBIAL PHYSIOLOGY-I

UNIT 1: MEMBRANE BIOENERGETICS

- 1.1 The Chemiosmotic theory
- 1.2 Electrochemical energy
- 1.3 Ionophores
- 1.4 Measurement of the Δp
- 1.5 Exergonic reactions that generate a Δp

UNIT 2: ELECTRON TRANSPORT

- 2.1 Aerobic and Anaerobic respiration
- 2.2 The electron carriers
- 2.3 Organization of electron carriers in bacteria and mitochondria
- 2.4 Coupling sites
- 2.5 Patterns of electron flow in *E. coli*

UNIT 3: METABOLITE TRANSPORT

- 3.1 Passive diffusion
- 3.2 Facilitated diffusion
- 3.3 Active transport
- 3.4 Group translocation
- 3.5 Iron uptake

UNIT 4: METABOLISM OF SUBSTRATES OTHER THAN GLUCOSE

- 4.1 Lactose
- 4.2 Galactose
- 4.3 Maltose
- 4.4 Mannitol
- 4.5 Fucose and Rhamnose

REFERENCES:

- 1. David White (2007). The Physiology and Biochemistry of Prokaryotes, 3rd Edition, Oxford University Press.
- 2. Moat, A. G., Foster, J. W. and Spector, M. P. (2002). Microbial Physiology, 4th Edition, John Wiley & Sons, USA.
- 3. Willey, J. M., Sherwood, L. M. and Woolverton, C. J. (2008). Prescott, Harley and Klein's Microbiology, 7th Edition, McGraw Hill International Edition.

VEER NARMAD SOUTH GUJARAT UNIVERSITY, SURAT M. Sc. Integrated Biotechnology Semester-III Syllabus: Practical: Core 1

Core-1; Course-1 Instrumentation & Techniques-I

- 1. Preparation of working solutions as well as different buffers and calibration of pH meter.
- 3. Study of Binocular Microscope and cell count by Haemocytometer.
- 4. To study the working of Centrifuge detailing analytical and preparative centrifugation.

Core-1; Course-2 Applied Microbiology

- 1. Detection of coliforms using P.A. test.
- 2. Enumeration of coliforms using MPN method.
- 3. Bacteriological analysis of milk using Methylene Blue Reduction Test.

Core-1; Course-3 Fundamentals of Immunology

- 1. Study of haemagglutination in blood grouping.
- 2. Study of bacterial agglutination using qualitative Widal slide test.
- 3. Study of precipitation reaction using quantitative single diffusion in two dimensions (Mancini Assay).

VEER NARMAD SOUTH GUJARAT UNIVERSITY, SURAT M. Sc. Integrated Biotechnology Semester-III Syllabus: Practical: Core 2

Core-2; Course-1 Plant Physiology-I

- 1. Determination of osmotic potential of cell sap by plasmolytic method
- 2. To study types of stomata based on the structure and number of subsidiary cells & to determine stomatal index and Frequency of the studied types.
- 3. To demonstrate that oxygen is evolved during photosynthesis & demonstration of plant movements.

Core-2; Course-2 Animal Physiology-I

- 1. Preparation of temporary mounts: Squamous epithelium, Ciliated epithelium, Striated muscle fibres and nerve cells.
- 2. Examination of sections of Mammalian skin, Cartilage, Bone, Pancreas, Testis, Ovary, Pituitary, Adrenal, Thyroid, Parathyroid.
- 3. Preparation of permanent slide of any five mammalian tissues- Microtomy.

Core-2; Course-3 Microbial Physiology-I

- 1. Carbohydrate Metabolism by *Escherichia coli*: Glucose, Lactose, Mannitol, Maltose, Sucrose and Xylose.
- 2. Carbohydrate Metabolism by *Bacillus subtilis*: Glucose, Lactose, Mannitol, Maltose, Sucrose and Xylose.
- 3. Carbohydrate Metabolism by *Pseudomonas aeruginosa*: Glucose, Lactose, Mannitol, Maltose, Sucrose and Xylose.

CORE: 1; Course: 01 INSTRUMENTATION & TECHNIQUES-II

Unit-I

- Spectroscopy:
 - Principle, instrumentation, interpretation and applications of IR spectroscopy
 - Working principle of NMR and Mass spectrometry and their applications
 - Atomic spectroscopy:
 - Flame emission spectroscopy: Working principle and applications
 - Atomic absorption spectroscopy principle, atomizers, source and applications
 - Inductively coupled plasma spectrometry: Principle, applications and advantages.

Unit-II

• Electrophoretic techniques:

- Principles of electrophoresis
- Support media
- Electrophoresis of proteins
 - $\circ \quad SDS PAGE$
 - Native gels
 - Gradient gels
 - o Isoelectric focusing
 - Two dimensional electrophoresis
 - o Western blotting
- Electrophoresis of Nucleic acids
 - Agarose gel of DNA, RNA
 - DNA sequencing gels
 - Southern blotting

Unit-III

• Chromatography:

- Classification of chromatography techniques
- Plane chromatography: Paper chromatography, TLC and HPTLC Methods of development –spot detection
- Principles of GC separation stationary and mobile phases
- \circ Detectors FID, TCD and ECD
- o Limitations of GC
- o Quantitative analysis and Qualitative analysis
- GC-MS combination

• Liquid chromatography:

- HPLC layout of instrument and components of instrumentation, Isocratic, binary and quaternary systems. Types of columns, stationary and mobile phases, detectors: UV absorption, PDA, RI and fluorescence,.
- Ion exchange chromatography: Types of Resins, principles of separation. Detection, applications.
- o Gel filtration chromatography: Principles of separation and applications

REFERENCES:

- 1. Keith Wilson & John Walker (Ed.) (2000): Practical Biochemistry Principles & Techniques. Cambridge University Press.
- 2. Skoog, Holler and Nieman, Instrumental analysis Saunders college publication
- 3. Skoog, West and Holler , Fundamentals of Analytical Chemistry Saunders college publication
- 4. James S. Fritz & George H. Schenk, Jr. (1969): Quantitative Analytical Chemistry (2nd edition). Allyn & Bacon, Inc., Boston.
- 5. Brown S.B. (1980): An Introduction to Spectroscopy for biochemists. Academic Press London.
- 6. Andrews A.T. (1986): Electrophoresis: Theory, Techniques and Biochemical and Clinical Application. Oxford University Press.
- 7. Hawcroft D.M. (1996): Electrophoresis The Basics. IRL Press, Oxford.
- 8. Robards K., Haddad P.R., & Jackson P.E. (1994): Principles and Practice of Modern Chromatographic Methods. Academic Press, London.

CORE: 1; Course: 02 ENZYMES & COENZYMES

UNIT 1: INTRODUCTION TO ENZYMES

- 1.1 Historical perspective
- 1.2 Substrate specificity
- 1.3 International classification of enzymes
- 1.4 Fisher 'lock-and-key' hypothesis
- 1.5 Koshland 'induced fit' hypothesis

UNIT 2: COENZYMES IN ENZYME CATALYZED REACTION

- 2.1 Nicotinamide Nucleotides (NAD⁺ and NADP⁺)
- 2.2 Flavin Nucleotides
- 2.3 Adenosine Phosphates (ATP, ADP and AMP)
- 2.4 Coenzyme A
- 2.5 Thiamine Pyrophosphate (TPP)

UNIT 3: ENZYMATIC REACTIONS

- 3.1 Chemical kinetics
- 3.2 Enzyme kinetics
- 3.3 Inhibition
- 3.4 Effects of pH
- 3.5 Bisubstrate reaction

UNIT 4: REGULATORY ENZYMES

- 4.1 Introduction
- 4.2 Role of non covalent binding of Modulators
- 4.3 Allosteric enzymes: as an exception to many general rules
- 4.4 Kinetic behaviour of Allosteric enzymes
- 4.5 Other mechanism of enzyme regulation.

REFERENCES:

- Lehninger, A., Cox, M. M., & Nelson, D.L., (2008). Principles of Biochemistry, 5th Edition, W. H. Freeman, New York.
- 2. Trevor Palmer (2004). Enzymes- Biochemistry, Biotechnology, Clinical chemistry, East-West Press Pvt Ltd.
- 3. Voet D. and Voet J. G. (2011). Biochemistry, 4th Edition, John-Wiley & Sons.

CORE: 1; Course: 03

INTRODUCTION TO MOLECULAR BIOLOGY

UNIT 1: CONCEPT OF GENOME AND ITS ORGANIZATION

- 1.1 History: Elucidation of DNA structure
- 1.2 DNA as genetic material
 - 1.2.1 Griffith's Transformation Experiment
 - 1.2.2 Hershey-Chase Experiment
- 1.3 Flow of genetic information
- 1.4 Genome organization: Structure of eukaryotic gene
- 1.5 Packaging of Viral, Bacterial and Eukaryotic DNA

UNIT 2: DNA REPLICATION

- 2.1 General features of DNA replication
- 2.2 Replication in Prokaryotes
- 2.3 Replication in Eukaryotes
- 2.4 Termination of Replication
- 2.5 Regulation of Replication

UNIT 3: TRANSCRIPTION

- 3.1 Establishment of Genetic code
- 3.2 Organization of Genetic code
- 3.3 Transcription in Bacteria
- 3.4 Transcription in Eukaryotes
- 3.5 Transcription in the Archaea

UNIT 4: TRANSLATION

- 4.1 Transfer RNA and Amino acid activation
- 4.2 The Ribosome
- 4.3 Initiation of Protein synthesis
- 4.4 Elongation of polypeptide chain
- 4.5 Termination of Protein synthesis

REFERENCES:

- Lehninger, A., Cox, M. M., & Nelson, D.L., (2008). Principles of Biochemistry, 5th Edition, W. H. Freeman, New York.
- 2. Willey, J. M., Sherwood, L. M. and Woolverton, C. J. (2008). Prescott, Harley and Klein's Microbiology, 7th Edition, McGraw Hill International Edition.
- 3. Pal, J. K. and Ghaskadbi, S. S. (2009). Fundamentals of Molecular Biology, Oxford University Press.

CORE: 2; Course: 01 PLANT PHYSIOLOGY-II

UNIT – 1 Metabolic Processes:

Structure of chloroplast; Significance of photosynthesis, photosynthetic pigments, action spectra, concept of two photosystem, photophosphorylation, C_3 , C_4 , CAM pathway; Photorespiration and its significance.

UNIT-2 Nitrogen Metabolism:

Nitrogen metabolism- Nitrogen cycle, Biology of Nitrogen fixation, Assimilation of Nitrate and Ammonium ions.

Lipid –Formation and metabolism. , Saturated and unsaturated fatty acids.

UNIT-3 Environmental Physiology:

Concept of biological clock and biorhythm.

Stress Physiology- Water stress; Drought, cold and salt- Physiological process affected by drought. Drought resistance mechanisms: Escape, dehydration postponement (Drought avoidance), Osmotic adjustment Osmoprotectants, stress proteins. High Temperature stress.

UNIT-4 Growth Regulation:

Physiological role of Phytohormones -auxins. Cytokinins, gibberellins, ABA and ethylene: their application. Methyl jasmonate, Brassinosteroids and Polyamines as PGRs (brief idea), Hydroponics and aeroponics. Cellular totipotency.

REFERENCES:

- 1. Frank B.Salisbury and Cleon W.Ross (2010), Plant Physiology, Cengage learning products, India Edition.
- 2. S.K. Verma and Mohit Verma (1999) Plant Physiology Biochemistry and Biotechnology, S Chand.
- 3. Lincoln Taiz and Eduardo Zaiger (Fourth Edition), Plant Physiology, Sinquur Associates inc. Publishers.
- 4. S.N. Pandey ajd K.K Singh, Plant physiology, Vikas Pub.
- 5. Medicinal Plant Biotechnology by Ciddi Veerasham. CBS Publishers.

CORE: 2; Course: 02 ANIMAL PHYSIOLOGY-II

UNIT-1 Respiratory System

Histology of trachea and lung; Pulmonary ventilation; Respiratory volumes and capacities; Transport of oxygen in the blood (oxygen-hemoglobin and myoglobin dissociation curve and its influencing factors), Carbon monoxide poisoning; Carbon dioxide transport in the blood; Regulation of acid-base balance; Control of respiration.

UNIT-2 Excretory System

Histology of kidney, ureter and bladder; Renal blood supply; Mechanism and regulation of urine formation; Regulation of acid-base balance; Renal failure and dialysis.

UNIT-3 Reproductive System

Histology of male and female reproductive systems, Puberty, physiology of male and female reproduction; Methods of contraception (depicted through flow chart); Disorders of reproductive system.

UNIT-4 Endocrine System

Histology and functions of endocrine glands; Nature of hormones; Regulation of hormone secretion; Mode of action of hormones, Hypothalamus- principal nuclei involved in control of endocrine system, control of anterior pituitary hormones by hypothalamic releasing hormones (neuroendocrine mechanisms); Effects of abnormal secretions of hormones; Placental hormones.

-----X------X-----

(Ch 28, Tortora & Grabowski)

(Ch 18, Tortora & Grabowski)

(Ch 23, Tortora & Grabowski)

(Ch 26, 27, Tortora & Grabowski)

CORE: 2; Course: 03 MICROBIAL PHYSIOLOGY-II

UNIT 1: CENTRAL METABOLIC PATHWAYS

- 1.1 Glycolysis
- 1.2 Pentose Phosphate Pathway
- 1.3 Entner-Doudoroff Pathway
- 1.4 Citric acid Cycle
- 1.5 Glyoxylate Cycle

UNIT 2: METABOLISM OF:

- 2.1 Lipids
- 2.2 Nucleotides
- 2.3 Amino acids
- 2.4 Aliphatic Hydrocarbons
- 2.5 CO₂

UNIT 3: BIOSYNTHESIS OF:

- 3.1 Peptidoglycan
- 3.2 Lipopolysaccharide
- 3.3 Dextran
- 3.4 Glycogen
- 3.5 Extracellular Polysaccharide synthesis

UNIT 4: FERMENTATIONS

- 4.1 Lactate
- 4.2 Butyrate
- 4.3 Mixed acid and Butanediol
- 4.4 Acetate
- 4.5 Propionate

REFERENCES:

- 1. David White (2007). The Physiology and Biochemistry of Prokaryotes, 3rd Edition, Oxford University Press.
- 2. Moat, A. G., Foster, J. W. and Spector, M. P. (2002). Microbial Physiology, 4th Edition, John Wiley & Sons, USA.
- Willey, J. M., Sherwood, L. M. and Woolverton, C. J. (2008). Prescott, Harley and Klein's Microbiology, 7th Edition, McGraw Hill International Edition.

17

VEER NARMAD SOUTH GUJARAT UNIVERSITY, SURAT M. Sc. Integrated Biotechnology Semester-IV Syllabus: Practical: Core 1

Core-1; Course- 1 Instruments & Techniques-II

- 1. To study the working of spectrophotometer and details of spectrophotometry.
- 2. Preparation of column and plates in Gel filtration and thin layer chromatography.
- 3. Preparation of Agarose and SDS polyacrylamide gels in electrophoresis.

Core-1; Course- 2 Enzymes & Coenzymes

- 1. Effect of pH on the rate of Acid Phosphatase Catalyzed reaction.
- 2. Effect of Temperature on the rate of Acid Phosphatase Catalyzed reaction.
- 3. Isoenzyme analysis: Acid Phospatase.

Core-1; Course- 3 Introduction to Molecular Biology

- 1. Isolation of Prokaryotic and Eukaryotic DNA.
- 2. Estimation of DNA by Diphenylamine method.
- 3. Estimation of RNA by Orcinol method.

VEER NARMAD SOUTH GUJARAT UNIVERSITY, SURAT M. Sc. Integrated Biotechnology Semester-IV Syllabus: Practical: Core 2

Core-2; Course-1 Plant Physiology-II

- 1. Extraction and separation of plant pigments (Chlorophylls and carotenoids) by chemical method through separating funnel.
- 2. Separation of anthocyanin & chlorophyll pigment by paper chromatography.
- 3. Identification of Common organic acids -
 - Oxalic acid
 - Malic acid
 - Citric acid
 - ➢ Tartaric acid

Core-2; Course-2 Animal Physiology-II

- 1. Estimation of haemoglobin using Sahli's haemoglobinometer.
- 2. Preparation of haemin and haemochromogen crystals.
- 3. Examination of sections of mammalian stomach, intestine, liver, spleen and kidney.

Core-2; Course-3 Microbial Physiology-II

- 1. Metabolism of Carbohydrate (Starch) by microorganism.
- 2. Metabolism of Protein (Casein) by microorganism.
- 3. Metabolism of Lipid (Tributyrene) by microorganism.