### Two Year M.Sc. (Zoology) Programme w.e.f. 2016–2017

### **Proposed Scheme for Choice Based Credit System**

### **SEMESTER - I**

	Course	Tea.	Credits	Evaluation		Marks
		Hrs		Internal	External	
1	Core (SB)	4	4	20	80	100
2	Core (ECB)	4	4	20	80	100
3	Core (IMM)	4	4	20	80	100
4	Core (TSFAI)	4	4	20	80	100
5	Practical (SB)	4	2	1	50	50
6	Practical (ECB)	4	2	1	50	50
7	Practical (IMM)	4	2	1	50	50
8	Practical (TSFAI)	4	2	-	50	50
	Total	32	24			600

Paper – I: Structural Biology [SB]

**Paper – II :** Environmental and Conservation Biology [ECB]

**Paper – III:** Immunology [IMM]

Paper – IV: Taxonomy, Systematics and Functional Anatomy of Invertebrates [TSFAI]

### Two Year M.Sc. (Zoology) Programme w.e.f. 2016–2017

### **Proposed Scheme for Choice Based Credit System**

### <u>SEMESTER – II</u>

Course		Tea. Hrs	Credits	Evaluation		Marks
				Internal	External	
1	Core (TTB)	4	4	20	80	100
2	Core (AP)	4	4	20	80	100
3	Core (MGDB)	4	4	20	80	100
4	Core (EFAV)	4	4	20	80	100
5	Practical (TTB)	4	2	-	50	50
6	Practical (AP)	4	2	-	50	50
7	Practical (MGDB)	4	2	-	50	50
8	Practical (EFAV)	4	2	-	50	50
	Total	32	24			600

**Paper – I :** Tools, Techniques and Biostatistics [TTB]

**Paper – II:** Animal Physiology [AP]

**Paper – III:** Molecular Genetics and Developmental Biology [MGDB]

**Paper – IV:** Evolution and Functional Anatomy of Vertebrates [EFAV]

# Semester I CORE PAPER

Paper I		Zoo_101	
Title	Structural Biology [SB]		
UNIT I–Ba	asic concepts of Biomolecules and Struct	ural Biology	15 Hrs
1.1		carbohydrates, proteins, amino acids, nuclei	c acids
1.2	Chemistry and structure of mono, oli and glycosides	go and polysaccharides. Deoxysugars, amino	sugars
1.3	Classification and structures of protein	ns – primary, secondary, tertiary and quaterna	ary.
1.4	Classification, structure and function cerebrosides, steroids	of lipids, fatty acids, triglycerides; phosph	olipds,
1.5	Nucleic acids – Structure of DNA and F	NA , DNA polymorphism, RNA types.	
UNIT II–E	Inzymes and Metabolism		15 Hrs
3.1	Classification, nomenclature and prop	erties of enzymes – catalysis and energy of –Menten Constant; (Km values) and LB plot; alation of enzyme activity	
3.2	Metabolism of carbohydrates – Glcoly oxidation; role of respiratory chain in	sis; TCA cycle; Gluconeogenesis; biological energy capture; ATP synthesis	
3.3		nation, deamination and decarboxylation	
3.4	Oxidation and biosynthesis of fatty ac	ds	
3.5	Metabolic disorders of different biome	olecules (carbohydrates, proteins, lipids)	
UNIT III–(	Cellular Organization		15 Hrs
2.1	Molecular organization and functions	of cell membranes	
2.2	Cell permeability – Transport across t Carrier proteins; Ion pumps; membra	he cell membrane; transport of small molecu ne bound enzymes	les;
2.3	Cell communications – Inter cellular of between the cells; strategies of chem	ommunication and gap junctions; chemical si ical signaling	gnaling
2.4	3 3	ceptors; signaling mediated cell surface recep P, G–proteins, Ca++, Inositol Triphosphate (IP <sub>3</sub>	
2.5	Cell cycle; molecular events in cell cyc	:le; mitotic spindle	
UNIT IV -	Functional Biology of Nucleic Acids		15 Hrs
4.1	DNA replication – semi conservative,	enzymology of DNA replication, replication of nation of replication process. Proof reading fu	
4.2	Enzymatic synthesis of RNA.		
4.3	Protein synthesis – Events of protein eukaryotes; post transcriptional proce	synthesis; transcription in prokaryotes and essing.	
4.4		s concept, translation in prokaryotes and euk	
4.5	DNA repair mechanism – High fidelity light, Eukaryotes repair systems.	of DNA sequence – Repair of damage caused	by UV

PRACTICAL	
1	Determination of proteins by Biuret method/ Folin Phenol method
2	Determination of glucose by Somogi / Anthrone method
3	Determination of lipids by Vanlin method
4	Determination of glycogen by Kemp's method
5	Estimation of cholesterol
6	Determination of enzyme activities of SDH and LDH
7	Effect of substrate concentration and pH on SDH activity
8	Protein fractionation using sodium sulphate
9	Estimation of DNA and RNA
10	Electrophoretic analysis of proteins/DNA
11	Feulgen reaction method for DNA localization.
12	Submission of assignment on structure of Biomolecules, mechanism of enzyme action ,
	Metabolic cycles, DNA, RNA, protein synthesis. [To be submitted at the time of
	Examination – 10 Marks]
Suggested E	Books
1	Textbook of Biochemistry by Harper
2	Textbook of Biochemistry by Lehninger
3	Textbook of Biochemistry by Stryer and Stryer
4	Textbook of Biochemistry by Conn and Stumpf
5	Textbook of Biochemistry by A.B.V. Rama Rao
6	Cell and molecular biology by De Robertis and De Robertis, 8 <sup>th</sup> ed.
7	Molecular Biology by Friefielder
8	Molecular cell biology by Darnell, Lodish and Baltimore (Scientific American Books)
9	Molecular biology by H. D. Kumar
10	Biochemistry and molecular biology by W. H. Elliot and D. C.Elliot(OU Press)
11	Molecular Biology of Cell by Bruce Alberts et al.
12	Cell by Karp
Syllabus Cor	mmittee
1	Prof. P. Nagaraja Rao
2	Dr. Rafath Yasmeen
3	Dr. B. Jyothi
4	Dr. S. Padmaja

### Semester I CORE PAPER

Paper II Title	Code Zoo_102 Environmental and Conservation Biology [ECB]	
	1 33	Hrs
1.1	Laws of limiting factor, Laws of minimum, Laws of Tolerance and Tragedy of commons	
1.2	Micronutrients and macronutrients	
1.3	Types of ecosystem – freshwater, marine and terrestrial	
1.4	Population characteristics and dynamics – conceptual approach	
1.5	Growth curves and pyramids; sigmoid curve, J curve and hyperbola; logistic equation a	nd
	concepts relating to growth	
UNIT II – C	Community Organization and Structure 15	Hrs
2.1	Community analysis, species diversity, ecotone concept and edge effect; interaction	
	between environment and biota Habitat and ecological niche and niche overlap; conce	ept
	of biome	
2.2	Concepts of productivity; eutrophication of lakes; biological indicator and water quality	y
2.3	Ecosystem dynamics and management; stability and complexity of ecosystem	
2.4	Biogeochemical cycles; inorganic pollutants and their impact SO <sub>2</sub> , NO <sub>2</sub> , CO, Phosphate	
	heavy metals (Arsenic, Lead and Mercury); radioactive nucleotides and their impact on	1
0.5	biological system	
2.5	Acid rain sources and its impact on biological system; green house effect and ozone	
	depletion	
UNIT III - E	Biogeography of India, Habitats and Resources 15	Hrs
3.1	Classical concepts of biogeography – continental drift, endemism, refugia	
3.2	Biogeographical regions of India and their salient features	
3.3	Classification, function and values of habitats – Freshwater wetlands, deserts, grasslan and forests	ds
3.4	Concepts of natural resources – renewable and non-renewable resources	
3.5	Overexploitation of resources – deforestation, water table depletion and land	
	degradation	
I INIT IV - N	Natural Resource Management 15	Hrs
4.1	Environmental Impact Assessment – principle, scope and purpose	1113
4.2	Role of ecological restoration in conservation; displacement and settlement of local	
	communities	
4.3	Major conservation movements in India; NGOs in conservation efforts	
4.4	Community diversity resources use and management; conflict management and	
	resolution	
4.5	National legislations for protecting biological resources – Biodiversity Act, 2002 and	
	Biodiversity Rules, 2004	

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- 1 Estimation of phosphates from the water sample
- 2 Estimation of nitrates and nitrites
- 3 Estimation of magnesium
- 4 Estimation of calcium
- Biological indicators of water quality and their population dynamics collection of water sample
- 6 Identification, enumeration of zooplankton, and their ecological significance
- 7 Estimation of total alkalinity of water and soil
- 8 Estimation of particulate matter in air
- 9 Draw the biogeographical regions of India and provide in brief the salient features of each biogeographical zone
- 10 Enumerate the biological diversity (zooplanktons and birds) using the habitat of freshwater lake in your place
- 11 Enumerate the diversity (plants and animals) use and their management in a community/village near your place

### Suggested Books

- Caughley, G., and A. Gunn. 1996. Conservation Biology in Theory and Practice. Blackwell Science, Cambridge, Massachusetts, U.S.A.
- 2 Cox, G. W. 2005. Conservation Biology: Concepts and Applications. McGraw-Hill, Dubuque, Iowa, U.S.A.
- 3 Dasmann, R., 1981. Wildlife Biology, 2nd ed. John Wiley and Sons, NY
- Dobson, A. P. 1996. Conservation and Biodiversity. Scientific American Library, New York, New York, U.S.A.
- Jeffries, M. J. 1997. Biodiversity and Conservation. Routledge, New York, New York, U.S.A.
- 6 Mills, L. Scott 2006. Conservation of Wildlife Populations. Blackwell Science, Oxford, U. K.
- Milner-Gulland, E. J., and R. Mace. 1998. Conservation of Biological Resources. Blackwell Science, Oxford, U.K.
- 8 Morris, W. F., and D. F. Doak2002. Quantitative Conservation Biology: Theory and Practice of Population Viability Analysis. Sinauer Associates, Sunderland, Massachusetts, U.S.A.
- 9 Sinclair, A. R. E., J. M. Fryxell, and G. Caughley2006. Wildlife Ecology, Conservation and Management, Blackwell Publishing
- Soulé ME (ed) 1986. Conservation biology: the science of scarcity and diversity- Sinauer, Sunderland
- Bram F. Noble 2005. Introduction to Environmental Impact Assessment: A Guide to Principles and Practice. Oxford University Press, London
- John A. Wiens and Michael R. Moss 2005. Issues and Perspectives in Landscape Ecology. Cambridge University Press, London
- Aparna Sawhney 2004. The New Face of Environmental Management in India. Ashgate Publishing Ltd., Sheffield

### Syllabus Committee

- 1 Prof. S. Jithender Kumar Naik
- 2 Dr. C. Srinivasulu
- 3 Dr. Apka Nageshwar Rao

# Semester I CORE PAPER

Paper III		Code Zoo_103
Title	Immunology [IM]	
UNIT I – Ir	ntroduction to Immune System	15 Hrs
1.1	Phylogeny of Immune system –invertebrate	s and vertebrates
1.2		unity , humoral mediated immunity and cell-
1.3	Cells involved in immune system; role of ma	crophages in immunity
1.4	The Lymphoid tissues – primary and second	ary lymphoid organs, lymphatic traffic
1.5	Activation of B– and T– Cells; production of	effectors – antibodies and cytokines
UNIT II – A	Antigen-Antibody Nature and Complement Sy	stem 15 Hrs
2.1	Antigens nature, epitope, haptens, antigen	presenting cells, adjuvants, antigenicity
2.2	Immunoglobulins structure, function and cla	assification of antibodies.
2.3	Monoclonal antibodies and its application. A	· ·
	techniques -Principles and applications of El GISH	·
2.4	Complement system – Components of comp	, ,
	alternative, biological consequences of com	plement activation and complement
	significance	
2.5	Major histocompatibility complex (MHC) str Immunoresponses; MHC restriction	ucture and function; genetic control of
UNIT III –	Hypersensitivity Reactions and Autoimmune	Diseases 15 Hrs
3.1	Hypersensitivity – Classification of hypersen hypersensitivity; Type – II Antibody – media	3, , , ,
3.2	Type-III – Immunocomplex mediated hyper hypersensitivity.	sensitivity; Type – IV Cell mediated (Delayed)
3.3	dependent diabetes mellitus (type-I diabete	•
3.4	Autoimmune diseases – Systemic autoimm Erythematosus (SLE), Rheumatoid arthritis.	une diseases – Systemic Lupus
3.5	Genetic factors, pathogenesis and treatmen	t of autoimmune diseases.
UNIT IV –	Transplantation and Tumour Immunology	15 Hrs
4.1	Transplantation – Barriers to transplantation	n.
4.2	Genetic predisposition for graft rejection, pr	
4.3	Immunity to infection – viruses, bacteria, fu immunopathological considerations.	
4.4	Tumor immunology – Immunity to tumors, t	tumor specific antigens.
4.5	Immunosurveillance.	

PRACTICAL
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1	Agglutination test
2	Precipitation
3	Demonstration of Immunoectrophoresis
1	Neutralization and complement fixation
5	Separation of lymphocytes
ó	Collection of macrophages and their characterization
7	Immunization schedules and rising of antibodies
3	Identification of histological slides of lymphoid tissue - Spleen, thymus, lymphnode and
	bone marrow
9	Demonstration of lymphocyte transformation test with nitrogen and an antigen

# Suggested Books

1	Immunology, Kuby, W.F.Freeman, U.S.A
2	Fundamentals of Immunology, W.Paul
3	Essentials of Immunology, I.M.Roitt
4	Immunology A Foundation Test by Basiro Davey
5	An introduction to immunology, by Ian R. Tizard

# Syllabus Committee

Prof. K. Pratap Reddy
Dr. G. Sunitha Devi
Dr. Rafath Yasmeen

### Semester I CORE PAPER

	CORE PAPER
Paper IV	Code Zoo_104
Title	Taxonomy, Systematics and Functional Anatomy of Invertebrates [TSFAI]
LINUT I D'	4511
	systematics and Taxonomy 15 Hrs
1.1	Basic concepts of biosystematics, taxonomy and classification
1.2	Recent trends in biosystematics ¬– molecular taxonomy
1.3	Taxonomic hierarchies, species concepts.
1.4	International Code for Zoological Nomenclature (ICZN) – operative principles,
	interpretation and application of important rules; basis of scientific names
1.5	Concepts of Prokarya, Eukarya, Protostomia and Deuterostomia; Significance of
	symmetry, coelom and metamerism
	A DI A LA L
	otozoa to Platyhelminthes 15 Hrs
2.1	Structure and functions of Locomotary organs in protozoans, reproduction in protozoa,
	conjugation in verticella
2.2	Porifera :-sycoin canal system, life cycle
2.3	Platyhelminthes:- Polyembryons
2.4	Life cycles and modes of transmission, Paragonimus westermani, Diphyllobothrium
	latum.
2.5	Overview of reproduction and development in Protozoa, Proifera, Cnidaria, Ctenophora
	and Platyhelminthes
LINIT III A.	nnelida to Echinodermata 15 Hrs
3.1	Filter feeding in polychaetes and respiration in Annelida.
3.2	Shell in mollusca; respiration in mollusca foot in molluscs
3.3	Arthropoda; social life in insects; respiration and excretion in arthropods
3.4	Echinodermata; Autotomy and regeneration in echinodumata
3.5	Overview of reproduction, development and phylogenetic significance of the larval forms
	of Arthropoda and Echinodermata
I INIT IV - M	linor and Other Phyla 15 Hrs
4.1	Systematic position, general organization and affinities of Ctenophora and Nemertea
7.1	(Rhynchoceola)
4.2	Systematic position, general organization and affinities of Rotifera
4.2	Systematic position, general organization and affinities of Rothera  Systematic position, general organization and affinities of Bryozoa (Ectoprocta).
4.3 4.4	
4.4	Systematic position, general organization and affinities of Onychophora and
1 E	Chaetognatha
	Nictomatic nacition, danaral arganization and attinition at Hamichardata
4.5	Systematic position, general organization and affinities of Hemichordata
4.0	Systematic position, general organization and affinities of Hemichordata
PRACTICAL	

Salient characteristics, identification and classification of representative types of invertebrate groups from Protozoa, Porifera, Cnidaria, Ctenophora, Annelida, Mollusca, Arthropoda, Echinodermata and Hemichordata

- 2 Preparation of permanent slides of mouth parts of house fly, mosquitoes and silk moth
- 3 Collection and identification of invertebrates in pond water
- 4 Collection and identification of parasites from cockroach
- 5 Dissections
  - 1. Minor a) Reproductive system of cockroach, b) Mouth parts of cockroach
  - 2. Major a) Nervous system of prawn

### Suggested Books

- 1 Principles of systematic Zoology (2 nd Edition) by E. Mayr and P.D. Ashlock
- 2 A Textbook of Zoology Vol. I by Parker and Haswell (Revised)
- The Invertebrates Vol. I to Vol. VI by L. H. Hyman
- 4 Invertebrate structure and function by E. J. W. Barrington
- 5 Invertebrate Zoology by P. A. Meglitsch (Oxford Press)
- 6. Life of Invertebrates by Russel Hunter
- 7. Invertebrate Zoology by Rupport and Barnes (Saunders College Publishing Co.)
- 8. Life of Invertebrates by S. N. Prasad
- 9. Evolutionary Biology by Eric C. Mitkoff
- 10. Worms and Man by D. W. T. Crompton
- 11. Parasitology by Noble and Noble
- 12. Regeneration by S. M. Rose–Appleton (Century Crofts)

### Syllabus Committee

- 1 Prof. Geeta Rajalingam
- 2 Dr. C. Srinivasulu
- 3 Dr. A. V. Rajashekar

### Semester II CORE PAPER

UNIT I - Tools and Separation Techniques 15 Hrs  1.1 Principles of microscopy – light, UV, confocal, phase contrast, fluorescent, electron microscopy (SEM & TEM)  1.2 Principles of histology – microtomy – tissue fixation, staining, mounting, histochemistry  1.3 Homogenization; cell fractionation; centrifugation – principles and applications of Preparative, analytical and ultra centrifugation – principles and applications of Preparative, analytical and ultra centrifugation permeation, ion-exchange, column, TLC, GLC, HPLC  1.5 Electrophoresis techniques – principles and applications of adsorption, affinity, partition, permeation, ion-exchange, column, TLC, GLC, HPLC  1.5 Spectroscopic techniques – principles and applications of visible, UV, fluorescence, IR, ESR, NMR and mass spectroscopy  2.2 Radioisotope techniques – principles and applications of visible, UV, fluorescence, IR, ESR, NMR and mass spectroscopy  2.3 Electrophysiological techniques – principles and applications of single neuron recording, patch clamp recording.  2.4 Imaging techniques – ECG, PET, MRI, fMRI and CAT  2.5 Microarray techniques – principles and applications of DNA, RNA and Protein microarray Techniques  1.5 Irs Statistical data, organization, classification and tabulation of data; Frequency distribution and graphical representation of data  3.2 Measures of central tendency – Mathematical average (Mean – Arithmetic, Geometric & Harmonic Mean) and Positional Averages (Median and Mode)  3.3 Measures of dispersion (or variability) – types, range, quartile deviation, mean deviation, variance, standard deviation, coefficient of variance  3.4 Basics of Probability – Concept of probability, addition and multiplication laws of probability and application to the problems of biology  3.5 Probability distribution – Definition, Types, properties and applications – Normal, Binomial and Poisson distributions  UNIT IV - Biostatistics II – Hypothesis testing and Inferential Statistics  4.1 Sampling – concept, sampling distribution of mean, standard err	Paper I	Zoo_201	
UNIT I - Tools and Separation Techniques  1.1 Principles of microscopy - light, UV, confocal, phase contrast, fluorescent, electron microscopy (SEM & TEM)  1.2 Principles of histology - microtomy - tissue fixation, staining, mounting, histochemistry  1.3 Homogenization; cell fractionation; centrifugation - principles and applications of Preparative, analytical and ultra centrifugation - principles and applications of Preparative, analytical and ultra centrifugation - principles and applications of adsorption, affinity, partition, permeation, ion-exchange, column, TLC, GLC, HPLC  1.5 Electrophoresis techniques - principle and applications of continuous, disc, isoelectrofocussing, isotachophoresis.  UNIT II - Separation and Diagnostic Techniques  2.1 Spectroscopic techniques - principles and applications of visible, UV, fluorescence, IR, ESR, NMR and mass spectroscopy  2.2 Radioisotope techniques - principles and application of Geiger-Muller counter, scintillation counter, tracer studies, autoradiography  2.3 Electrophysiological techniques - principles and applications of single neuron recording, patch clamp recording.  2.4 Imaging techniques - ECG, PET, MRI, fMRI and CAT  2.5 Microarray techniques - principles and applications of DNA, RNA and Protein microarray Techniques  UNIT III - Biostatistics I - Introduction, Measures and Theories of Probability  15 Hrs  3.1 Statistical data, organization, classification and tabulation of data; Frequency distribution and graphical representation of data  3.2 Measures of central tendency - Mathematical average (Mean - Arithmetic, Geometric & Harmonic Mean) and Positional Averages (Median and Mode)  3.3 Measures of central tendency - Mathematical average (Mean - Arithmetic, Geometric & Harmonic Mean) and Positional Averages (Median and Mode)  3.4 Basics of Probability - Concept of probability, addition and multiplication laws of probability and application to the problems of biology  3.5 Probability and application to the problems of biology  3.5 Probability - Concept of pr	•	——————————————————————————————————————	
<ul> <li>Principles of microscopy – light, UV, confocal, phase contrast, fluorescent, electron microscopy (SEM &amp; TEM)</li> <li>Principles of histology – microtomy – tissue fixation, staining, mounting, histochemistry Homogenization; cell fractionation; centrifugation – principles and applications of Preparative, analytical and ultra centrifugation</li> <li>Chromatographic techniques – principle and applications of adsorption, affinity, partition, permeation, ion-exchange, column, TLC, GLC, HPLC</li> <li>Electrophoresis techniques – principles and applications of continuous, disc, isoelectrofocussing, isotachophoresis.</li> <li>UNIT II - Separation and Diagnostic Techniques</li> <li>Spectroscopic techniques – principles and applications of visible, UV, fluorescence, IR, ESR, NMR and mass spectroscopy</li> <li>Radioisotope techniques – principles and application of Geiger-Muller counter, scintillation counter, tracer studies, autoradiography</li> <li>Electrophysiological techniques – principles and applications of single neuron recording, patch clamp recording.</li> <li>Imaging techniques - EGG, PET, MRI, fMRI and CAT</li> <li>Microarray techniques – principles and applications of DNA, RNA and Protein microarray Techniques</li> <li>UNIT III - Biostatistics I – Introduction, Measures and Theories of Probability</li> <li>Statistical data, organization, classification and tabulation of data; Frequency distribution and graphical representation of data</li> <li>Measures of central tendency – Mathematical average (Mean – Arithmetic, Geometric &amp; Harmonic Mean) and Positional Averages (Median and Mode)</li> <li>Measures of dispersion (or variability) – types, range, quartile deviation, mean deviation, variance, standard deviation, coefficient of variance</li> <li>Basics of Probability – Concept of probability, addition and multiplication laws of probability and application to the problems of biology</li> <li>Probability dropability, addition and multiplicati</li></ul>	TILLE	1001s, recrimques and biostatistics [11b]	
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<ul> <li>Principles of microscopy – light, UV, confocal, phase contrast, fluorescent, electron microscopy (SEM &amp; TEM)</li> <li>Principles of histology – microtomy – tissue fixation, staining, mounting, histochemistry Homogenization; cell fractionation; centrifugation – principles and applications of Preparative, analytical and ultra centrifugation</li> <li>Chromatographic techniques – principle and applications of adsorption, affinity, partition, permeation, ion-exchange, column, TLC, GLC, HPLC</li> <li>Electrophoresis techniques – principles and applications of continuous, disc, isoelectrofocussing, isotachophoresis.</li> <li>UNIT II - Separation and Diagnostic Techniques</li> <li>Spectroscopic techniques – principles and applications of visible, UV, fluorescence, IR, ESR, NMR and mass spectroscopy</li> <li>Radioisotope techniques – principles and application of Geiger-Muller counter, scintillation counter, tracer studies, autoradiography</li> <li>Electrophysiological techniques – principles and applications of single neuron recording, patch clamp recording.</li> <li>Imaging techniques - EGG, PET, MRI, fMRI and CAT</li> <li>Microarray techniques – principles and applications of DNA, RNA and Protein microarray Techniques</li> <li>Statistical data, organization, classification and tabulation of data; Frequency distribution and graphical representation of data</li> <li>Measures of central tendency – Mathematical average (Mean – Arithmetic, Geometric &amp; Harmonic Mean) and Positional Averages (Median and Mode)</li> <li>Measures of dispersion (or variability) – types, range, quartile deviation, mean deviation, variance, standard deviation, coefficient of variance</li> <li>Basics of Probability – Concept of probability, addition and multiplication laws of probability and application to the problems of biology</li> <li>Probability distribution – Definition, Types, properties and applications – Normal, Binomial and Poisson distributions</li> <li>UNIT IV - B</li></ul>	UNIT I - To	pols and Separation Techniques 15	Hrs
microscopy (SEM & TEM)  1.2 Principles of histology – microtomy – tissue fixation, staining, mounting, histochemistry  1.3 Homogenization: cell fractionation; centrifugation – principles and applications of Preparative, analytical and ultra centrifugation  1.4 Chromatographic techniques – principle and applications of adsorption, affinity, partition, permeation, ion-exchange, column, TLC, GLC, HPLC  1.5 Electrophoresis techniques – principles and applications of continuous, disc, isoelectrofocussing, isotachophoresis.  UNIT II - Separation and Diagnostic Techniques 15 Hrs  2.1 Spectroscopic techniques – principles and applications of visible, UV, fluorescence, IR, ESR, NMR and mass spectroscopy  2.2 Radioisotope techniques – principles and application of Geiger-Muller counter, scintillation counter, tracer studies, autoradiography  2.3 Electrophysiological techniques – principles and applications of single neuron recording, patch clamp recording.  2.4 Imaging techniques - ECG, PET, MRI, fMRI and CAT  2.5 Microarray techniques – principles and applications of DNA, RNA and Protein microarray Techniques  UNIT III - Biostatistics I – Introduction, Measures and Theories of Probability 15 Hrs  3.1 Statistical data, organization, classification and tabulation of data; Frequency distribution and graphical representation of data  3.2 Measures of central tendency – Mathematical average (Mean – Arithmetic, Geometric & Harmonic Mean) and Positional Averages (Median and Mode)  3.3 Measures of dispersion (or variability) – types, range, quartile deviation, mean deviation, variance, standard deviation, coefficient of variance  3.4 Basics of Probability – Concept of probability, addition and multiplication laws of probability and application to the problems of biology  3.5 Probability and application to the problems of biology  3.5 Probability distribution – Definition, Types, properties and applications – Normal, Binomial and Poisson distributions  UNIT IV - Biostatistics II – Hypothesis testing and Inferential Statistics  4.		·	
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31 . 31	1.4		1
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4.3	Nonparametric tests – Assumptions, applications of Sign Test, Wilcoxon Signed Rank Test, Man-Whitney Test
4.4	Parametric tests – Student's t-Test; Analysis of Variance (ANOVA or F-Ratio: One way and Two-way analysis); Chi-square test (Test of Independence and Test of Goodness of Fit)
4.5	Correlation and regression analysis concepts and their application
PRACTICAL	
1	To fix a tissue with Bouin's fixative and stain using haemotoxylin – eosin stain for histochemical studies
2	To prepare a paraffin block of tissue for microtomy for making sections of tissue for histochemical studies
3	Quantitative detection of total carbohydrates using Anthrone technique
4	Quantitative detection of total lipids using sulpho-phospovanillin technique
5	Quantitative detection of total proteins using Lowry et al's Biuret technique
6	Graphic presentation of data – bar diagram, histogram, frequency polygon and pie chart
7	Calculation of measures of central tendencies – mean, median and mode
8	Calculation of measures of dispersions – range, mean deviation, standard deviation, variance and coefficient of variance
9	Computation of test of significance – comparison of sample mean with population mean and two sample means
10	Calculate the coefficient of correlation between two variables
11	Computation of linear regression
12	Computation of One Way Analysis of Variance (ANOVA)
13	Using Chi Square Test, test the independence of two variables
Suggested E	Books
1	Principles and techniques of Practical Biochemistry Ed. B.L. Williams & D. Wilson, Arnold Publishers
2	Practical Biochemistry By Plummer
3	Immunology – Roit
4	Cell and Molecular Biology – DeRoberties
5	Cell and Molecular Biology – Ladish et al.
6	Statistical methods, Snedecor, G.W. and W.G. Cochran, Iowa State Univ. Press
7	Biometry by W. H. Freeman and Francisco
8	Fundamentals of Biometry by L.N. Balaram (1980)
9	Biostatistics by N. Gurumani
10	Techniques in life sciences – by Tembhare

Syllabus Committee 1 Prof. B. Reddya Naik 2 Dr. C. Srinivasulu

### Semester II CORE PAPER

	CORE PAPER	
Paper II		Code Zoo_202
Title	Animal Physiology [AP]	
11110	7 tillitar i rigorology [7 ti ]	
LIMIT I D	gestion-Respiration- Circulation	15 Hrs
	•	
1.1	Cellulose digestion –Ruminant and non-ruminant digestion; ab	
	events of absorptive and post absorptive states and their regul	ation (endocrine and
	neural).	•
1 2	•	itudo: adaptation to
1.2	Respiration – Cascade of oxygen transport to tissues at high alt	itude; adaptation to
	diving.	
1.3	Responses to $CO_2$ and $O_2$ rich environment; oxygen toxicity; hy	percapnea, control of
	respiration.	' '
1 /	·	
1.4	Buffering mechanisms by body fluids.	
1.5	Circulation - Cardiac cycle and principles of hemodynamics; blo	ood
	coagulation, haematome formation; Anti-coagulants.	
	3	
LINIT II– O	smoregulation, Excretion and thermoregulation	15 Hrs
2.1	Osmoregulation – Osmoregulatory problems in brackish water	
	organisms; osmotic problems in terrestrial animals; hormonal of	control of osmoregulation.
2.2	Excretion – Urine formation, counter current mechanism; juxta	nglomerular apparatus,
	rennin-angiotensin system; hormonal regulation – ADH and alc	losterone
2.3		
2.3	Detoxification of nitrogen products; purine cycle and miscellan	eous detoxilication
	pathways.	
2.4	Thermal physiology – temperature regulation in poikilotherms,	homeotherms and
	heterotherms, and their mechanisms of survival; central control	ol of homeothermy.
2.5	Cold death, cold resistance, heat death; Torpor, hibernation an	3
2.5	cold death, cold resistance, heat death, for por, hisemation an	id acstivation.
LINIT III A	Augala Dhyaialagu Nauranhyaialagu ( Dagantara	1F Uro
	Muscle Physiology, Neurophysiology & Receptors	15 Hrs
3.1	Comparative molecular structure and function of skeletal, smo	oth and cardiac muscles;
	energy metabolism in skeletal muscle, muscle fatigue.	
3.2	Types of neurons and glial cells.	
3.3	Basis and significance of membrane potentials, equilibrium pot	tontials their change
3.3		termais, men change
	during stimulus, Na, K currents in action potential.	
3.4	Types of synapses, synaptic transmission - electrical and chemi	cal; synaptic inhibition and
	neurotransmitters.	
3.5	Receptors – Receptor mechanisms, sensory coding; Mechanore	acentors photochemical
5.5	·	eceptors, priotocrientical
	aspects of vision and phonoreception in mammals.	
UNIT IV – I	Endocrinology, Bioluminiscence & Stress Physiology	15 Hrs
4.1	Structure and function of endocrine glands of invertebrate.	
4.2	Structure and function of endocrine glands of vertebrate.	
	· · · · · · · · · · · · · · · · · · ·	`
4.3	Mechanism of hormone action (peptide and steroid hormones)	
4.4	Bioluminiscence-luminiscent organisms-neural control. Biocher	mistry and significance of
	luminoscopeo	

luminescence.

4.5	Stress – resistance to stress, functions of hormones and sympathetic nervous system in stress
PRACTICAL	
1	Estimation of blood chlorides under hetero osmotic media.
2	Cold and heat stress on metabolic rate in tilapia fish/crab.
3	Effect of heat stress on glycogen levels in tilapia fish/crab.
4	Estimation of Acetylcholinesterase activity.
5	Estimation of phosphorylase activity.
6	Adrenalin and insulin induced changes in blood glucose levels in rat/mice.
7	Kymographic recordings of twitch, tetanus and fatigue.
8	Estimation of Hb, ESR and blood clotting time.
9	Cell fragility.

### Suggested Books

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1	Principles of Animal Physiology by D.W. Wood.
2	Principles of Animal Physiology by Gordon.
3	Animal Physiology-Adaptations and environment by Schmidt-Nielson.
4	Principles of Animal Physiology by Wilson.
5	Text Book of Medical Physiology by Guyton.
6	General & Comparative Animal Physiology By William Hoar.
7	Comparative Animal Physiology by Florey.
8	Comparative Animal Physiology by L.C.Prosser.
9	Human Physiology by vander .

# Syllabus Committee

Prof. Sugita Mathur
 Dr. Rafath Yasmeen
 Dr. S. Padmaja
 Dr. Jyothi

### Semester II CORE PAPER

Paper III	Code Zoo_203
Title	Molecular Genetics and Developmental Biology [MGDB]
UNIT I - In	troduction to Genetics 15 Hrs
1.1	Mendelism, mendelian inheritance; modification of mendelian inheritance.
1.2	Linkage studies, crossing over and extra chromosomal inheritance, multiple alleles, blood group antigens.
1.3	Chromosome structure (Prokaryote and Eukaryote); identification, karyotype.
1.4	Genetic disorders – chromosomal disorders, inborn errors of metabolism and polygenic and environmental disorders.
1.5	Bacterial genetics – transformation, transduction, conjugation, viral lytic and lysogenic cycle.
UNIT II – N	Molecular Genetics 15 Hrs
2.1	Introduction of DNA technology – Restriction endonucleases, methods of ligation – DNA ligases, ligation of fragment with cohesive and blunt ends.
2.2	Features of vectors – cosmids, plasmids and shuttle vector with one example representing each class construction and characterization of new cloning vectors
2.3	Applied molecular biology – DNA sequences – Maxam and Gilbert methods, Sanger's method. Application of recombinant DNA technology with reference to the example of insulin, somatostatin, and interferon. DNA fingerprinting and its application
2.4	Cloning strategies – Shotgun cloning, construction of gene libraries, genomic library and DNA library
2.5	Hybridization techniques – Southern blot, Northern blot, R-loop mapping methods, Insitu hybridization
UNIT III –	Overview of Developmental Biology 15 Hrs
3.1	Scope and importance of developmental biology
3.2	Gametogenesis; spermatogenesis, oogenesis, vitellogenesis and chemodifferentiation
3.3	Fertilization, parthenogenesis and its significance
3.4	Types of cleavage, mechanism of cleavage, chemical changes during cleavage
3.5	Role of cytoplasm and nucleus during early development; morphogenetic movements, presumptive areas and fate maps
UNIT IV –	Organogenesis 15 Hrs
4.1	Gastrulation, metabolic events during gastrulation and rudimentary organs formation
4.2	Concept of organisers and induction – Neural tubule formation
4.3	Organogenesis: limb, central nervous system, heart, kidney and eye
4.4	Role of hormones in metamorphosis of insects and frog; regeneration in Cnidaria, Echinodermata, Amphibia (limb and tail regeneration), and Reptiles (tail regeneration)
4.5	Teratogenesis– genetic and environmental; developmental mechanisms of teratogenesis

PRACTICAL	
1	Identification of ABO Blood groups
2	Extraction of DNA from tissues
3	Extraction and isolation of RNA from tissues
4	Estimation of RNA, DNA in tissues
5	Estimation of structural proteins
6	Estimation of soluble proteins
7	Estimation of SDH activity in chick embryo
8	Estimation of LDH activity in chick embryo
9	Estimation of calcium in egg shell by EDTA method
10	Identification of chick embryo developmental stages – 24hrs, 48hrs, 72hrs, &96hrs
Suggested E	Books
1	General genetics by Winchester
2	Molecular Biology of gene by Watson et al. Vol I & II
3	Genetics by Strickberger
4	Molecular Biology by Friefielder
5	Genetics by P.K. Gupta
6	Genes by Lewis
7	General genetics by S. R. B. Owen
8	Cell and molecular biology by De Robertis and De Robertis, 8th ed.
11	Molecular cell biology by Darnell, Lodish and Baltimore (Scientific American books)
12	Molecular biology by H. D. Kumar
13	Biochemistry and molecular biology by W. H. Elliot and D. C.Elliot (OUPress)
14	Text book of molecular biology by K. S. Sastry et al. (MacMillan Ind. Pvt. Ltd.)
15	Developmental Biology - patterns, problems and principles by W. Saunders Jr.
16	Principles of Animal Developmental Biology by S.C. Goel
17	Introduction to embryology by Balinsky
18	Developmental Biology S. Gilbert
19	Evolution by Savage
20	Process of organic evolution by Stebbings

# Syllabus Committee

21 22

Prof. V. Vanitha Das
 Dr. G. Sunitha Devi
 Dr. A.V. Rajashekar

Evolution of vertebrates by Colbert Developmental Biology by Berryl

# Semester II CORE PAPER

Evolution and Functional Anatomy of Vertebrates [EFAV]

Code Zoo 204

Paper IV

Title

UNIT I – Evolution 15 Hrs 1.1 Concept of evolution and theories of evolution 1.2 Variation, gene mutation and chromosomal aberrations in evolution; genetic drift 1.3 Speciation – species concepts, categories; Modes of speciation – Allopatric, parapatric and sympatric speciation 1.4 Natural selection; patterns of evolution – sequential, divergent, convergent, gradual, punctuated, monophyletic, polyphyletic and paraphyletic 1.5 Origin and evolution of primates and human UNIT II – Evolution of Vertebrates 15 Hrs Origin and salient features of Ostracoderm, Placoderm, Acanthodii, Sarcopterygii and 2.1 Actinopterygii 2.2 Origin, salient features and adaptive radiation in amphibians – Lepospondyli and Lissamphibia 2.3 Origin, salient features and adaptive radiation in early and Mesozoic reptiles 2.4 Origin, salient features and adaptive radiation in birds – Palaeognathae and Neognathae 2.5 Origin, salient features and adaptive radiation in mammals – Prototheria and Theriiformes UNIT III – Functional Anatomy of Vertebrates – from fishes to mammals 15 Hrs 3.1 Integumentary system ¬– Integument and its derivatives 3.2 Skeletal system ¬– Cranial and Post-Cranial (axial and appendicular) skeletal system 3.3 Nervous system – brain, spinal cord and peripheral nerves; sense organs 3.4 Respiratory and circulatory system; Digestive and excretory system 3.5 Reproductive system – comparison of male and female reproductive systems from fishes to mammals UNIT IV – Functional Anatomy of Vertebrates – Evolutionary significance 15 Hrs 4.1 Evolutionary significance of internal fertilization, neoteny and paedogenesis 4.2 Amniotic egg ¬– structure and its evolutionary significance 4.3 Basic plan of skull; Temporal fossae and their evolutionary significance; Vertebrate Jaw suspension 4.4 Types and evolutionary significance of axial and appendicular joints 4.5 Types and evolutionary significance of placenta; evolutionary significance of opposable thumb and bipedalism in primates (both non-human and human)

### PRACTICAL

- Salient characteristics, identification and classification of representative types of vertebrate groups from Pisces, Amphibia, Reptilia, Aves and Mammalia
- 2 Collection and preparation of slides of ticks, mites, bed bug, human lice, fleas, mosquitoes and house flies
- 3 Structure, bionomics and biology of earthworms. Commercially important prawns, mussels and pearl oysters, harmful and useful insects and moths, cultivable fishes and frogs
- 4 Dissections ¬–
  - 1. Minor a) Weberian ossicles of Labeo, and b) Respiratory trees of Clarius
  - 2. Major a) a) Cranial nerves of Labeo (V, VII, IX & X cranial nerves, b) Cornea and pectin of chick

### Suggested Books

- 1 Principles of systematic Zoology (2 nd Edition) by E. Mayr and P.D. Ashlock.
- 2 A Textbook of Zoology Vol. I by Parker & Haswell (Revised)
- The Invertebrates Vol. I to Vol. VI by L. H. Hyman
- 4 Invertebrate Structure and Function by E. J. W. Barrington.
- 5 Invertebrate Zoology by P. A. Meglitsch (Oxford Press)
- 6 Life of Invertebrates by Russel Hunter
- 7 Invertebrate Zoology by Rupport and Barnes (Saunders College Publishing Co.).
- 8 Life of Invertebrates by S. N. Prasad
- 9 Evolutionary Biology by Eric C. Mitkoff
- 10 Worms and Man by D. W. T. Crompton
- 11 Regeneration by S. M. Rose–Appleton (Century Crofts).
- 12 Parasitology by Noble & Doble.

### Syllabus Committee

- 1 Prof. V. Vanitha Das
- 2 Dr. C. Srinivasulu
- 3 Dr. B. Neeraja

### Two Year M.Sc. (Zoology) Programme w.e.f. 2016–2017

### **Proposed Scheme for Choice Based Credit System**

### **SEMESTER - I**

	Course Tea.		Credits	Evaluation		Marks
		Hrs		Internal	External	
1	Core (SB)	4	4	20	80	100
2	Core (ECB)	4	4	20	80	100
3	Core (IMM)	4	4	20	80	100
4	Core (TSFAI)	4	4	20	80	100
5	Practical (SB)	4	2	1	50	50
6	Practical (ECB)	4	2	1	50	50
7	Practical (IMM)	4	2	-	50	50
8	Practical (TSFAI)	4	2	-	50	50
	Total	32	24			600

Paper – I: Structural Biology [SB]

**Paper – II :** Environmental and Conservation Biology [ECB]

**Paper – III:** Immunology [IMM]

Paper – IV: Taxonomy, Systematics and Functional Anatomy of Invertebrates [TSFAI]

### Two Year M.Sc. (Zoology) Programme w.e.f. 2016–2017

### **Proposed Scheme for Choice Based Credit System**

### <u>SEMESTER – II</u>

Course		Tea. Hrs	Credits	Evaluation		Marks
				Internal	External	
1	Core (TTB)	4	4	20	80	100
2	Core (AP)	4	4	20	80	100
3	Core (MGDB)	4	4	20	80	100
4	Core (EFAV)	4	4	20	80	100
5	Practical (TTB)	4	2	-	50	50
6	Practical (AP)	4	2	-	50	50
7	Practical (MGDB)	4	2	-	50	50
8	Practical (EFAV)	4	2	-	50	50
	Total	32	24		1	600

**Paper – I :** Tools, Techniques and Biostatistics [TTB]

**Paper – II:** Animal Physiology [AP]

**Paper – III:** Molecular Genetics and Developmental Biology [MGDB]

**Paper – IV:** Evolution and Functional Anatomy of Vertebrates [EFAV]